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(FILE 'HOME' ENTERED AT 08:08:40 ON 25 SEP 2001)
      FILE 'MEDLINE' ENTERED AT 08:08:52 ON 25 SEP 2001
  L1
              427 S POOLED SAMPLE#
  L2
                2 S L1 AND STOOL
 L3
            8861 S PATIENT POPULATION
 1.4
              427 S POOLED SAMPLE#
 L5
                0 S L3 AND L4
 L6
           21985 S P53
 L7
            9124 S ENERGY TRANSFER
 L8
              22 S L6 AND L7
 L9
          230226 S MUTATION#
 L10
              12 S L8 AND L9
 L11
            1314 S P53 AND (POINT (W) MUTATION)
 L12
               2 S ENERGY RESONANCE TRANSFER
 L13
               0 S L11 AND RHODAMINE
 L14
               0 S L11 AND FLUORESCEIN
 L15
             611 S L11 AND DETECT?
 L16
              0 S L11 AND PRIMER EXTENSION
 L17
              36 S L15 AND PROBE#
 L18
              92 S L15 AND PRIMER#
 L19
              8 S L17 AND RAS
 L20
              22 S P53 AND (ENERGY TRANSFER)
 L21
              11 S P53 AND RHODAMINE
 L22
              1 S L21 AND FLUORESCEIN
 L23
              0 S P53 AND FRET
 L24
              4 S RAS AND FRET
 L25
               1 S POOL(4A) PATIENT(4A) SAMPLE?
 L26
               6 S POOL? (4A) STOOL (4A) SAMPLE?
     FILE 'CAPLUS' ENTERED AT 08:28:47 ON 25 SEP 2001
L27
              3 S L26
L28
              4 S L23
L29
              0 S L5
L30
              57 S POOL? AND STOOL?
L31
              0 S MUTATION AND L30
L32
              1 S DNA AND L30
L33
            681 S FRET
           1261 S FLUORESCEN? (2A) ENERGY (2A) RESONANCE (2A) TRANSFER
L34
L35
           1417 S L33 OR L34
L36
              1 S L35 AND (STOOL OR EXCRET?)
L37
              7 S L35 AND POOL?
L38
           4634 S POPULATION AND SCREEN?
L39
              2 S L35 AND L38
L40
            516 S POPULATION (2A) SCREEN?
L41
            187 S MUTATION? AND L40
        1127941 S SAMPLE?
L42
L43
             43 S L41 AND L42
             41 S L40 AND (POOL? OR COMBINE OR COMBINING OR COMBINED)
L44
L45
             10 S L44 AND SAMPLE?
L46
           1436 S POOL? (2A) SAMPLE?
L47
             0 S L46 AND FRET
L48
             4 S L46 AND STOOL
L49
             0 S L48 AND DNA
L50
             0 S L48 AND MUTATION
L51
            10 S L45
L52
            4 S L48
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L28 ANSWER 4 OF 4 CAPLUS COPYRIGHT 2001 ACS
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AN 1998:307053 CAPLUS

DN 129:37221

TI Detection of point mutation using double fluorescent-labeled probes and detection of gene abnormalities by the method

IN Hirano, Kenichi

PA Hamamatsu Photonics K. K., Japan

SO Jpn. Kokai Tokkyo Koho, 14 pp.

CODEN: JKXXAF

DT Patent

LA Japanese

FAN.CNT 1

PΙ

AB Point mutation in a specific sequence of target nucleic acids is detected by hybridizing the target nucleic acids with a complementary probe labeled

with a fluorescent dye at one end and another fluorescent dye with the other end, both dyes show fluorescence resonance energy transfer (FRET) between them, at a higher temp. than room temp. and measuring the ratio of fluorescence intensity of one fluorescent dye to that of the other dye at the max. absorption wavelengths. Gene abnormalities, e.g. of oncogenes, are detected by the above method. FRET efficacy is dependent on temp. and fluorescence intensity is measured in various temp. points. For example, mutations in the gene p53 of liver samples were detected.

The diagnosis of old and new gastrointestinal parasites.

AU Long E G; Christie J D

CS Division of Bacterial and Mycotic Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia, USA.

SO CLINICS IN LABORATORY MEDICINE, (1995 Jun) 15 (2) 307-31. Ref: 140 Journal code: DLS; 8100174. ISSN: 0272-2712.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)
General Review; (REVIEW)
(REVIEW, TUTORIAL)

LA English

FS Priority Journals

EM 199510

ED Entered STN: 19951026 Last Updated on STN: 19951026 Entered Medline: 19951019

AB For the foreseeable future, light microscopy will continue to be the method of choice for diagnosing gastrointestinal parasites. However, in selected circumstances, the use of commercially available immunofluorescent kits will supersede the use of routine light microscopy for diagnosis of Giardia lamblia and Cryptosporidium parvum. These techniques may used to diagnose invasive amebic infections caused by E. histolytica in the future. Pooling stool

samples from the same or even different patients may offer a means
to process specimens in a more efficient and cost-effective manner
without

lowering the predictive value of an ova and parasite examination. Although

we suggest that, with some exceptions, stools for ova and parasite examination should not be accepted past the fourth day of hospitalization,

we cannot recommend the use of a single stool sample for diagnosis without

extensive studies in individual parasitology laboratories. Techniques have

still not been developed for the optimum methods of concentration of stool

for diagnosis of coccidian infections. For most laboratories, the diagnosis of microsporidian infections remains problematic because of the lack of a commercial source for oocysts to provide positive control material. (Note: There is now a commercial source for oocysts available.)

- 45 ANSWER 6 OF 10 CAPLUS COPYRIGHT 2001 ACS
- AN 1999:402039 CAPLUS
- DN 131:40257
- TI Applications of constant denaturant capillary electrophoresis/high-fidelity polymerase chain reaction to human genetic analysis
- AU Li-Sucholeiki, Xiao-Cheng; Khrapko, Konstantin; Andre, Paulo C.; Marcelino, Luisa A.; Karger, Barry L.; Thilly, William G.
- CS Division Bioengineering Environmental Health, Center Environmental Health Sciences, Massachusetts Institute Technology, Cambridge, MA, 02142, USA
- SO Electrophoresis (1999), 20(6), 1224-1232 CODEN: ELCTDN; ISSN: 0173-0835
- PB Wiley-VCH Verlag GmbH
- DT Journal; General Review
- LA English
- AΒ A review is given on the author's own works with 32 refs. Const. denaturant capillary electrophoresis (CDCE) permits high-resoln. sepn. of single-base variations occurring in an 100 bp isomelting DNA sequence based on their differential melting temps. By coupling CDCE for highly efficient enrichment of mutants with high-fidelity PCR (hifi PCR), the authors developed an anal. approach to detecting point mutations at frequencies .gtoreq.10(tm) in human genomic DNA. The authors present several applications of this approach in human genetic studies. authors have measured the point mutational spectra of a 100 bp mitochondrial DNA sequence in human tissues and cultured cells. The observations have led to the conclusion that the primary causes of mutation in human mitochondrial DNA are spontaneous in origin. In the course of studying the mitochondrial somatic mutations, the authors have also identified several nuclear pseudogenes homologous to the analyzed mitochondrial DNA fragment. Recently, through developments of the means to isolate the desired target sequences from bulk genomic DNA and to increase the loading capacity of CDCE, the authors have extended the CDCE/hifi PCR approach to study a chem. induced mutational spectrum in a single-copy nuclear sequence. Future applications of the CDCE/hifi PCR approach to human genetic anal. include studies of somatic mitochondrial mutations with respect to aging, measurement of mutational spectra of nuclear genes in healthy human tissues and population screening for disease-assocd. single nucleotide polymorphisms

(SNPs) in large pooled samples.

RE.CNT 32

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ΑN
     91183169
                 MEDLINE
DN
     91183169 PubMed ID: 2009369
     Mutation of the p53 gene in human acute myelogenous leukemia.
ΤI
     Slingerland J M; Minden M D; Benchimol S
CS
     Ontario Cancer Institute, Toronto, Canada.
SO
     BLOOD, (1991 Apr 1) 77 (7) 1500-7.
     Journal code: A8G; 7603509. ISSN: 0006-4971.
CY
     United States
     Journal; Article; (JOURNAL ARTICLE)
DT
LΑ
     English
     Abridged Index Medicus Journals; Priority Journals
     199105
ED
     Entered STN: 19910526
     Last Updated on STN: 19970203
     Entered Medline: 19910506
     Heterogeneity of {\bf p53} protein expression is seen in blast cells
AΒ
    of patients with acute myelogenous leukemia (AML). p53 protein
     is detected in the blasts of certain AML patients but not in
     others. We have identified p53 protein variants with abnormal
     mobility on gel electrophoresis and/or prolonged half-life (t 1/2). We
    have sequenced the {\bf p53} coding sequence from primary blast cells
    of five AML patients and from the AML cell line (OCIM2). In OCIM2, a
    point mutation in codon 274 was identified that changes
    a valine residue to aspartic acid. A wild type p53 allele was
     not detected in these cells. Two point mutations (codon 135,
    cysteine to serine; codon 246, methionine to valine) were identified in
    cDNA from blasts of one AML patient. Both mutations were present in blast
    colonies grown from single blast progenitor cells, indicating that
    individual leukemia cells had sustained mutation of both p53
    alleles. The cDNAs sequenced from blast samples of four other patients,
    including one with prolonged p53 protein t 1/2 and one with no
    detectable p53 protein, were fully wild type. Thus, the
    heterogeneity of p53 expression cannot be explained in all cases
    by genetic change in the p53 coding sequence. The prolonged t
    1/2 of p53 protein seen in some AML blasts may therefore reflect
    changes not inherent to p53. A model is proposed in which
    mutational inactivation of p53, although not required for the
    evolution of neoplasia, would confer a selective advantage, favoring
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MEDLINE

clonal outgrowth during disease progression.

L17 ANSWER 35 OF 36

- N 94031784 PubMed ID: 8217795
- TI Occurrence of point mutations in p53 gene is not increased in patients with acute myeloid leukaemia carrying an activating N-ras mutation.
- AU Buhler-Leclerc M; Gratwohl A; Senn H P
- CS Institut fur Medizinische Mikrobiologie, Universitat Basel, Switzerland.
- SO BRITISH JOURNAL OF HAEMATOLOGY, (1993 Jul) 84 (3) 443-50. Journal code: AXC; 0372544. ISSN: 0007-1048.
- CY ENGLAND: United Kingdom
- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 199312
- ED Entered STN: 19940117 Last Updated on STN: 19940117 Entered Medline: 19931207
- AB The frequency of simultaneously **detecting** N-ras and p53 gene mutations was studied in leukaemia cells of patients with acute myeloid leukaemia (AML) or with myelodysplastic syndrome (MDS). Using in vitro DNA amplification followed by oligonucleotide hybridization

analysis, 45 AML and six MDS patients were screened for activating mutations in codons 12, 13 and 61 of N-ras. Ten of them (eight AML and two MDS) were found positive. These 10 patients and 10 others without activating N-ras mutation were further analysed by direct sequencing of the amplified exons for p53 mutations and for atypical N-ras mutations. Beside the activating mutations in the N-ras gene, no additional transforming or nontransforming mutations could be detected in the N-ras. However, exon 7 of p53 was mutated in two AML patients without activating N-ras mutation. These data show that p53 mutations occurred with half the frequency of N-ras mutations in AML and that no positive correlation could be found between the onset of mutations

in N-ras and p53 genes.